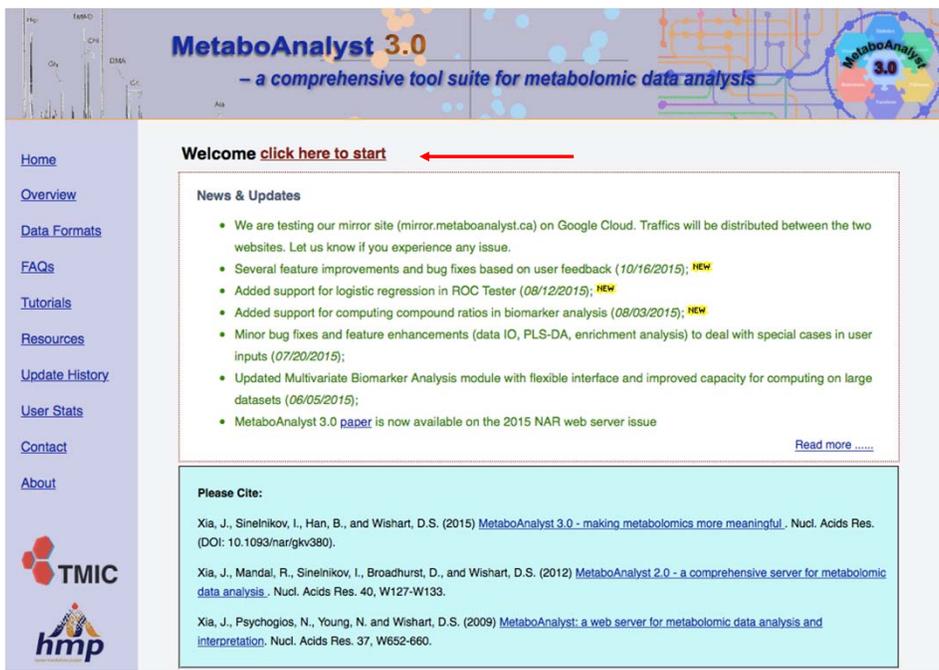


# Applying MetaboAnalyst

Xiangqin Cui, PhD



  
**MetaboAnalyst 3.0**  
 – a comprehensive tool suite for metabolomic data analysis

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**News & Updates**

- We are testing our mirror site ([mirror.metaboanalyst.ca](#)) on Google Cloud. Traffics will be distributed between the two websites. Let us know if you experience any issue.
- Several feature improvements and bug fixes based on user feedback (10/16/2015); **NEW**
- Added support for logistic regression in ROC Tester (08/12/2015); **NEW**
- Added support for computing compound ratios in biomarker analysis (08/03/2015); **NEW**
- Minor bug fixes and feature enhancements (data IO, PLS-DA, enrichment analysis) to deal with special cases in user inputs (07/20/2015);
- Updated Multivariate Biomarker Analysis module with flexible interface and improved capacity for computing on large datasets (06/05/2015);
- MetaboAnalyst 3.0 [paper](#) is now available on the 2015 NAR web server issue

[Read more .....](#)

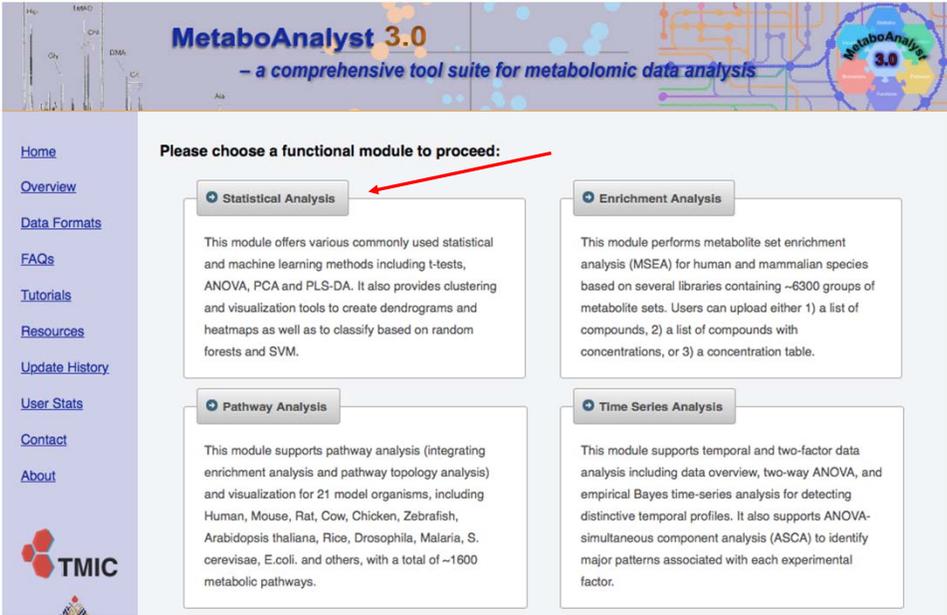
**Please Cite:**

Xia, J., Sinenikov, I., Han, B., and Wishart, D.S. (2015) [MetaboAnalyst 3.0 - making metabolomics more meaningful](#). Nucl. Acids Res. (DOI: 10.1093/nar/gkv380).

Xia, J., Mandal, R., Sinenikov, I., Broadhurst, D., and Wishart, D.S. (2012) [MetaboAnalyst 2.0 - a comprehensive server for metabolomic data analysis](#). Nucl. Acids Res. 40, W127-W133.

Xia, J., Psychogios, N., Young, N. and Wishart, D.S. (2009) [MetaboAnalyst: a web server for metabolomic data analysis and interpretation](#). Nucl. Acids Res. 37, W652-660.

**MetaboAnalyst 3.0**  
– a comprehensive tool suite for metabolomic data analysis

Please choose a functional module to proceed:

- Statistical Analysis** (indicated by a red arrow)
 

This module offers various commonly used statistical and machine learning methods including t-tests, ANOVA, PCA and PLS-DA. It also provides clustering and visualization tools to create dendrograms and heatmaps as well as to classify based on random forests and SVM.
- Enrichment Analysis**

This module performs metabolite set enrichment analysis (MSEA) for human and mammalian species based on several libraries containing ~6300 groups of metabolite sets. Users can upload either 1) a list of compounds, 2) a list of compounds with concentrations, or 3) a concentration table.
- Pathway Analysis**

This module supports pathway analysis (integrating enrichment analysis and pathway topology analysis) and visualization for 21 model organisms, including Human, Mouse, Rat, Cow, Chicken, Zebrafish, Arabidopsis thaliana, Rice, Drosophila, Malaria, S. cerevisiae, E.coli. and others, with a total of ~1600 metabolic pathways.
- Time Series Analysis**

This module supports temporal and two-factor data analysis including data overview, two-way ANOVA, and empirical Bayes time-series analysis for detecting distinctive temporal profiles. It also supports ANOVA-simultaneous component analysis (ASCA) to identify major patterns associated with each experimental factor.

Navigation links: Home, Overview, Data Formats, FAQs, Tutorials, Resources, Update History, User Stats, Contact, About

TMIC logo

**1) Upload your data**

**Tab-delimited text (.txt) or comma-separated values (.csv) file:**

Data Type:  Concentrations  Spectral bins  Peak intensity table

Format:

Data File:  No file selected.

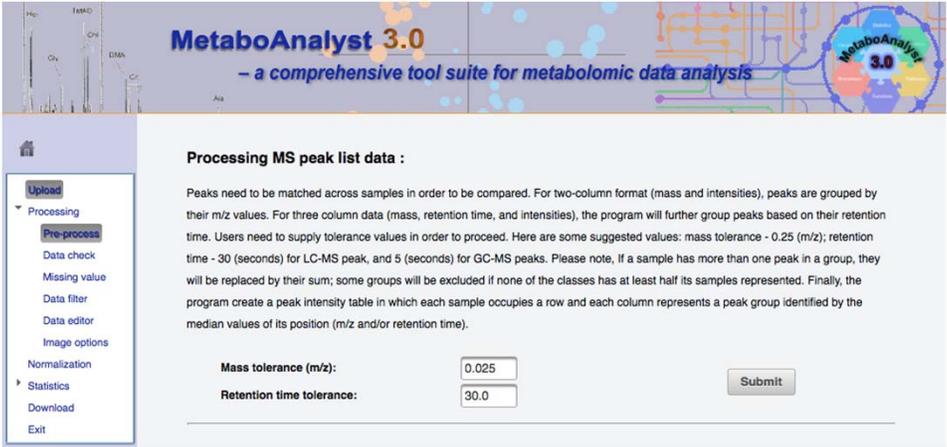
**Zipped Files (.zip) :**

Data Type:  NMR peak list  MS peak list  MS spectra (indicated by a red arrow)

Data File:  Posmode\_diet.zip (indicated by a red arrow)

Pair File:  No file selected.

**Select MS peak list option and then load the .zip file**



**MetaboAnalyst 3.0**  
– a comprehensive tool suite for metabolomic data analysis

**Processing MS peak list data :**

Peaks need to be matched across samples in order to be compared. For two-column format (mass and intensities), peaks are grouped by their m/z values. For three column data (mass, retention time, and intensities), the program will further group peaks based on their retention time. Users need to supply tolerance values in order to proceed. Here are some suggested values: mass tolerance - 0.25 (m/z); retention time - 30 (seconds) for LC-MS peak, and 5 (seconds) for GC-MS peaks. Please note, if a sample has more than one peak in a group, they will be replaced by their sum; some groups will be excluded if none of the classes has at least half its samples represented. Finally, the program create a peak intensity table in which each sample occupies a row and each column represents a peak group identified by the median values of its position (m/z and/or retention time).

Mass tolerance (m/z):

Retention time tolerance:

**Processing MS peak list data :**

Peaks need to be matched across samples in order to be compared. For two-column format (mass and intensities), peaks are grouped by their m/z values. For three column data (mass, retention time, and intensities), the program will further group peaks based on their retention time. Users need to supply tolerance values in order to proceed. Here are some suggested values: mass tolerance - 0.25 (m/z); retention time - 30 (seconds) for LC-MS peak, and 5 (seconds) for GC-MS peaks. Please note, if a sample has more than one peak in a group, they will be replaced by their sum; some groups will be excluded if none of the classes has at least half its samples represented. Finally, the program create a peak intensity table in which each sample occupies a row and each column represents a peak group identified by the median values of its position (m/z and/or retention time).

Mass tolerance (m/z):

Retention time tolerance:

---

**MS peak processing information**

The uploaded files are peak lists and intensities data.

A total of 6 samples were found.

These samples contain a total of 14304 peaks.

with an average of 2384 peaks per sample

A total of 2346 peak groups were formed.

Peaks of the same group were summed if they are from one sample.

Peaks appear in less than half of samples in each group were ignored.

**Data Integrity Check:**

1. Checking the class labels - at least three replicates are required in each class.
2. If the samples are paired, the pair labels must conform to the specified format.
3. The data (except class labels) must not contain non-numeric values.
4. The presence of missing values or features with constant values (i.e. all zeros)

**Data processing information:**

Checking data content ...passed

The uploaded files are peak lists and intensities data.

A total of 6 samples were found.

These samples contain a total of 14304 peaks.

with an average of 2384 peaks per sample

2 groups were detected in samples.

Samples are not paired.

All data values are numeric.

A total of 0 (0%) missing values were detected.

By default, these values will be replaced by a small value.

Click **Skip** button if you accept the default practice

Or click **Missing value imputation** to use other methods

Missing value estimation

Skip

**Note that XCMSonline filled in peaks**

Non-informative variables can be characterized in two groups: variables of very small values - these variables can be detected using mean or median; variables that are near-constant throughout the experiment conditions - these variables can be detected using standard deviation (SD); or the robust estimate such as interquartile range (IQR). The relative standard deviation( $RSD = SD/mean$ ) is another useful variance measure independent of the mean. The following empirical rules are applied during data filtering:

- **Less than 250 variables:** 5% will be filtered;
- **Between 250 - 500 variables:** 10% will be filtered;
- **Between 500 - 1000 variables:** 25% will be filtered;
- **Over 1000 variables:** 40% will be filtered;

Please note, in order to reduce the computational burden to the server, the **None** option is only for less than 2000 features. Over that, if you choose None, the IQR filter will still be applied. In addition, the maximum allowed number of variables is 5000. If over 5000 variables were left after filtering, only the top 5000 will be used in the subsequent analysis.

- Interquartile range (IQR)
- Standard deviation (SD)
- Median absolute deviation (MAD)
- Relative standard deviation ( $RSD = SD/mean$ )
- Non-parametric relative standard deviation (MAD/median)
- Mean intensity value
- Median intensity value
- None (less than 2000 features)

Process

**Sample normalization**

None

Sample specific normalization (i.e. dry weight, volume) [Click here to specify](#)

Normalization by sum

Normalization by median

Normalization by reference sample

Specify a reference sample

Create a pooled average sample from group

Normalization by reference feature

## Data options before stats analysis

**Data transformation**

None

Log transformation (generalized logarithm transformation or glog)

Cube root transformation (take cube root of data values)

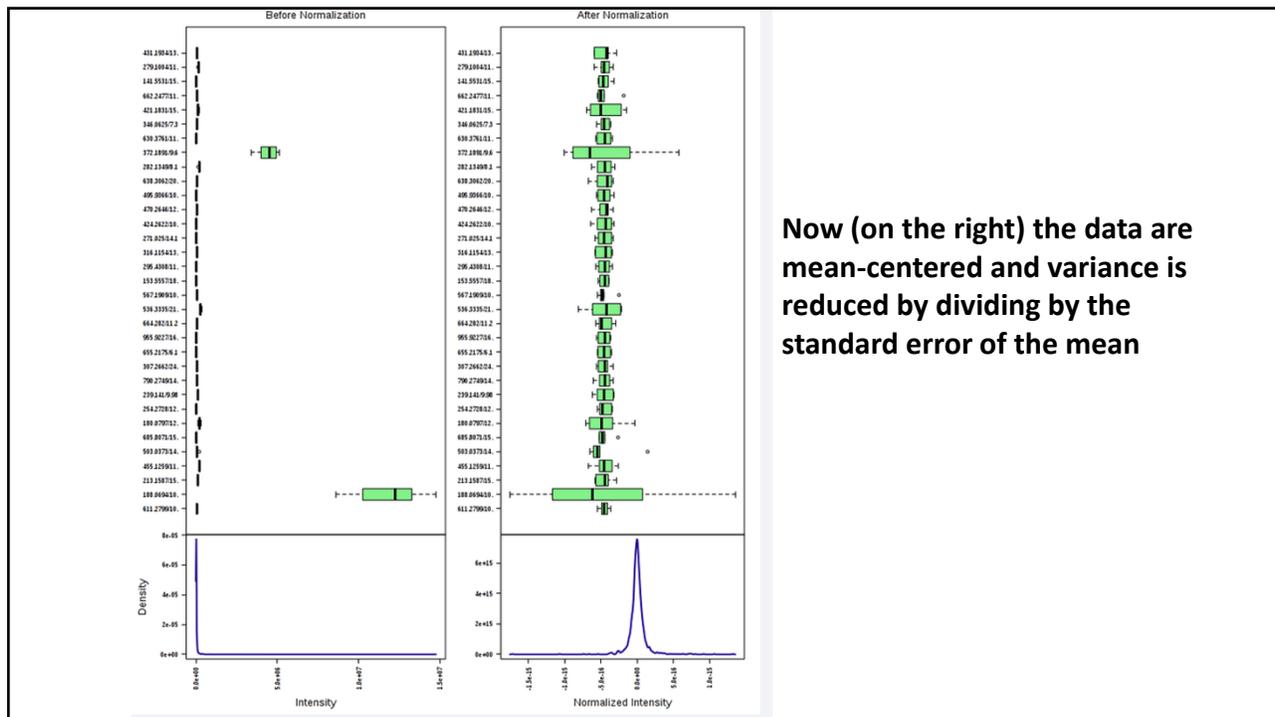
**Data scaling**

None

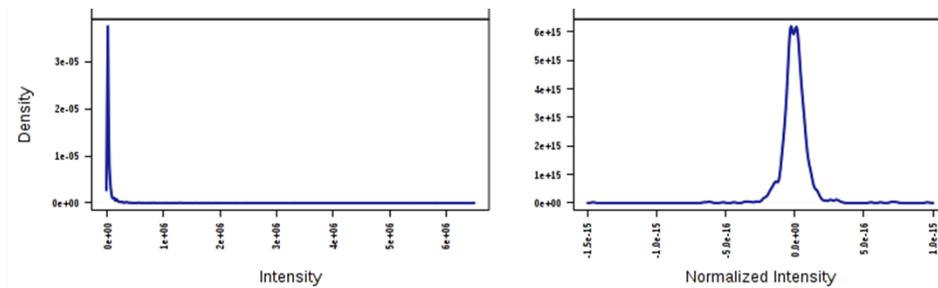
Auto scaling (mean-centered and divided by the standard deviation of each variable)

Pareto scaling (mean-centered and divided by the square root of standard deviation of each variable)

Range scaling (mean-centered and divided by the range of each variable)



## Effect of normalization, mean centering and Pareto scaling



**Univariate Analysis**[Fold Change Analysis](#) [T-tests](#) [Volcano plot](#)

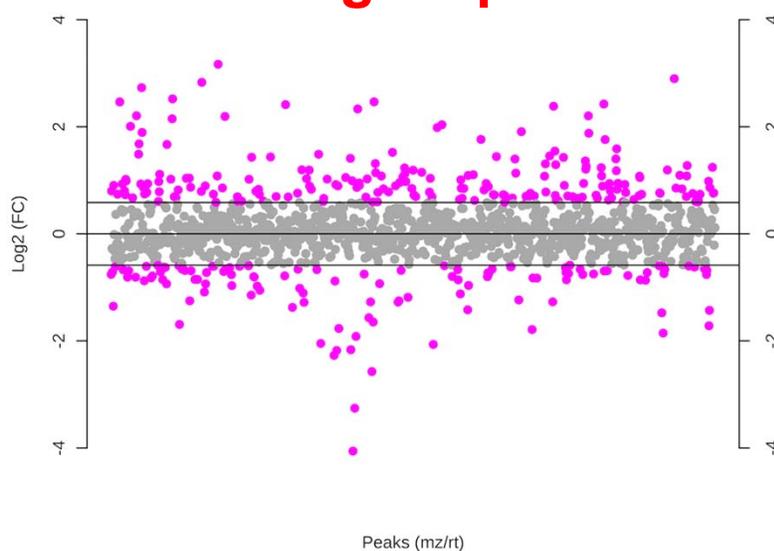
One-way Analysis of Variance (ANOVA)

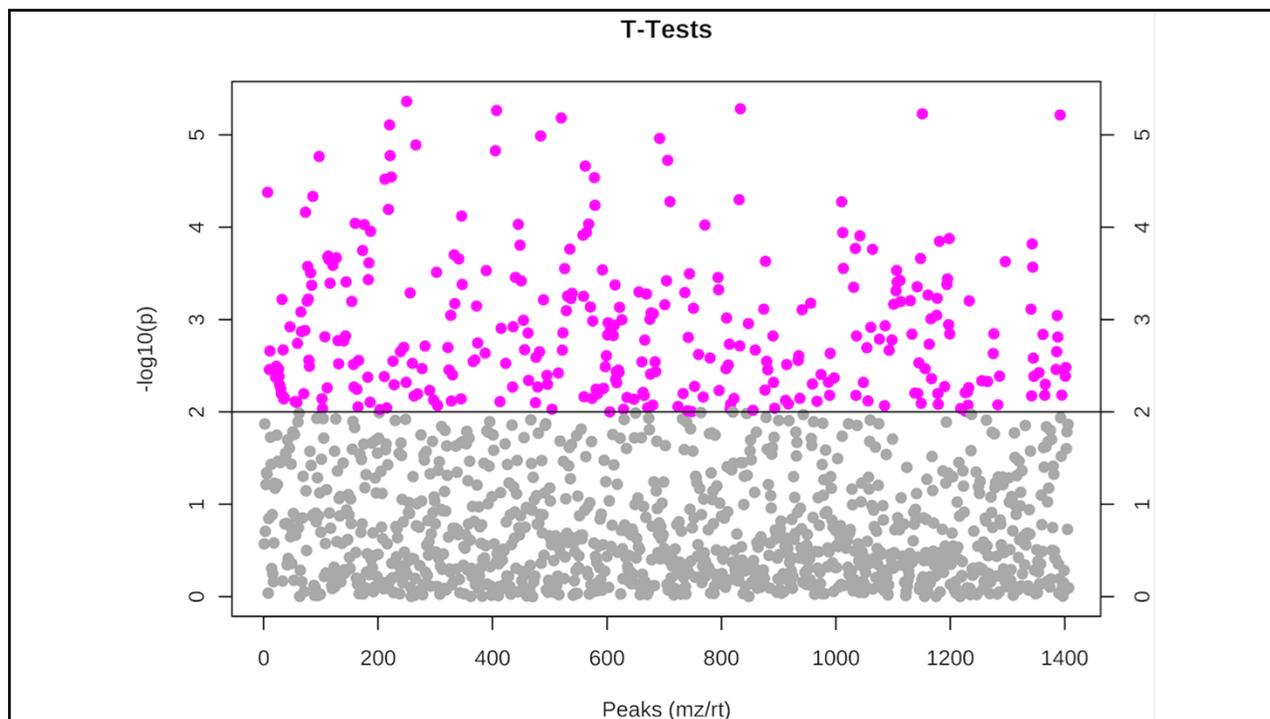
[Correlation Analysis](#) [Pattern Searching](#)**Multivariate Analysis**[Principal Component Analysis \(PCA\)](#)[Partial Least Squares - Discriminant Analysis \(PLS-DA\)](#)**Significant Feature Identification**[Significance Analysis of Microarray \(and Metabolites\) \(SAM\)](#)[Empirical Bayesian Analysis of Microarray \(and Metabolites\) \(EBAM\)](#)**Cluster Analysis**Hierarchical Clustering: [Dendrogram](#) [Heatmaps](#)Partitional Clustering: [K-means](#) [Self Organizing Map \(SOM\)](#)**Classification & Feature Selection**[Random Forest](#)[Support Vector Machine \(SVM\)](#)

Statistical methods available to process the data on MetaboAnalyst

Today we'll focus on univariate analysis (Volcano plots) and multivariate analysis (PCA and PLS-DA)

**Fold-change - pink = >1.5**





## Univariate analysis – Volcano plot

- This is a useful measure
- The data points represent comparisons made one metabolite at a time
- The plot identifies metabolites with p-values  $<0.01$  and a fold change  $>1.5$

## Volcano plot set up

**Volcano Plot**

The volcano plot is a combination of fold change and t-tests. Note, for unpaired samples, the x-axis is log (FC). For paired analysis, the x-axis is number of significant counts. Y-axis is  $-\log_{10}(\text{p.value})$  for both cases.

Analysis type:

Fold change threshold:

X-axis: Comparison type:

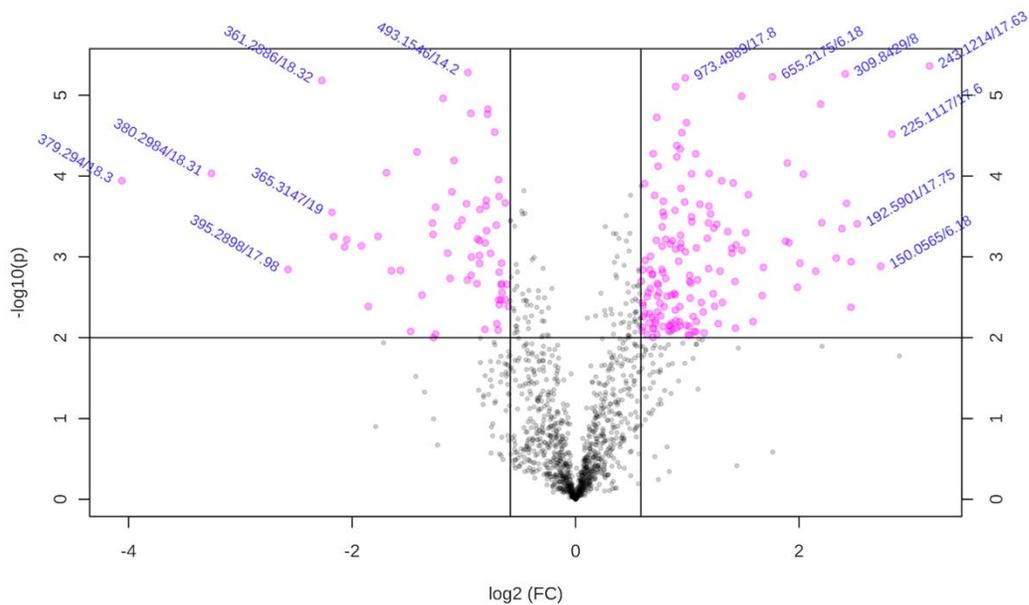
Sig. count threshold (paired):  %

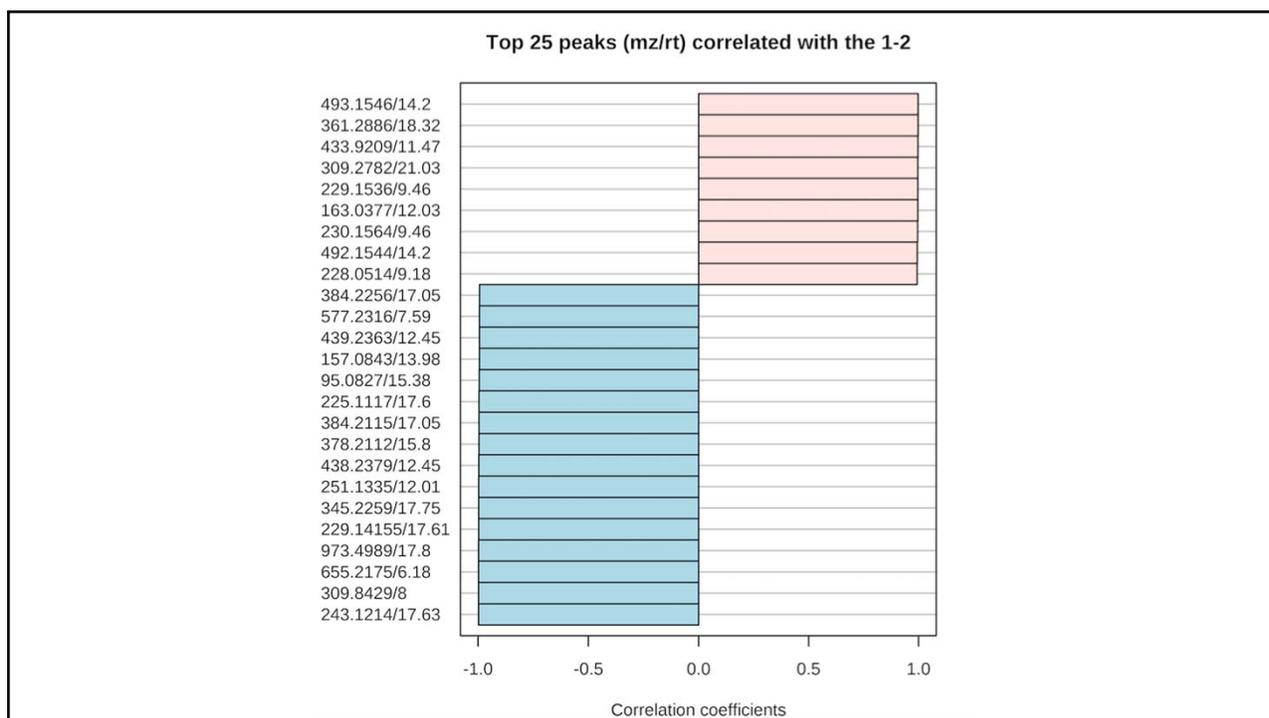
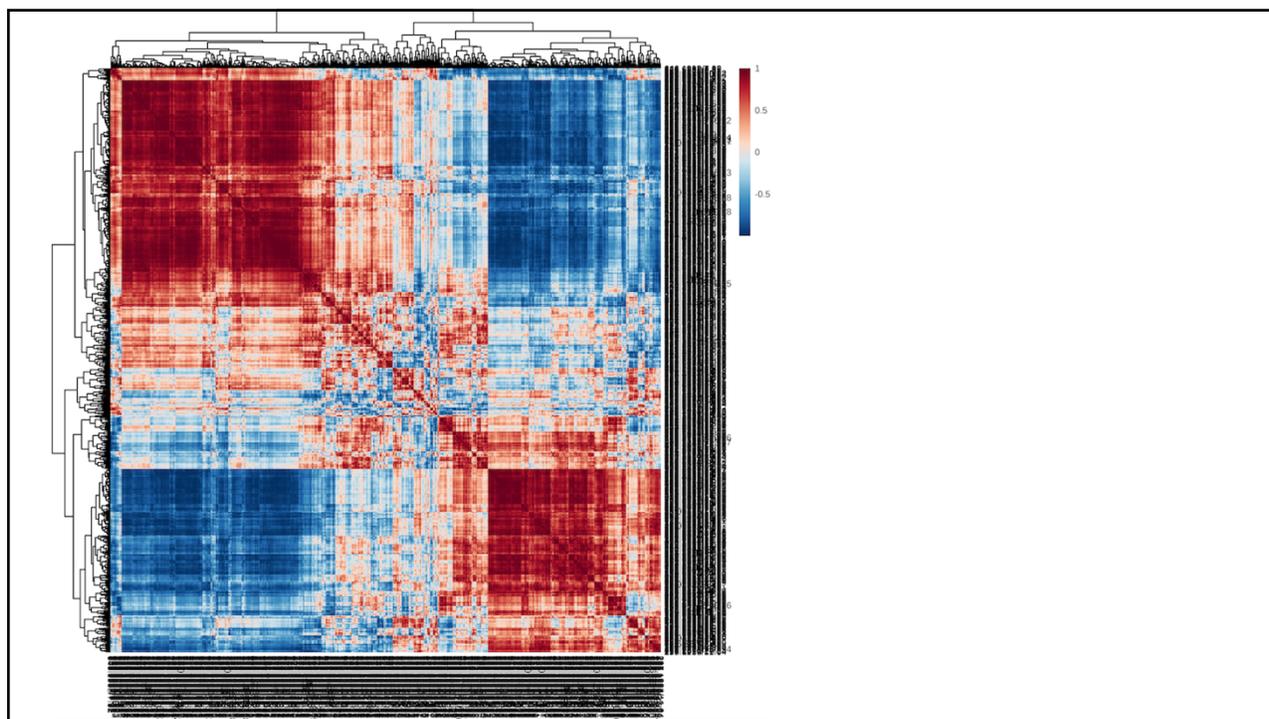
Non-parametric tests:

Y-axis: P value threshold:

Group variance:

## Volcano plot with fold change=1.5 and $p < 0.01$

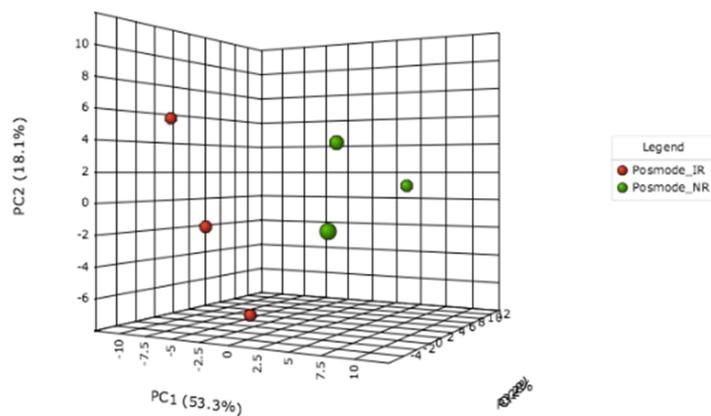
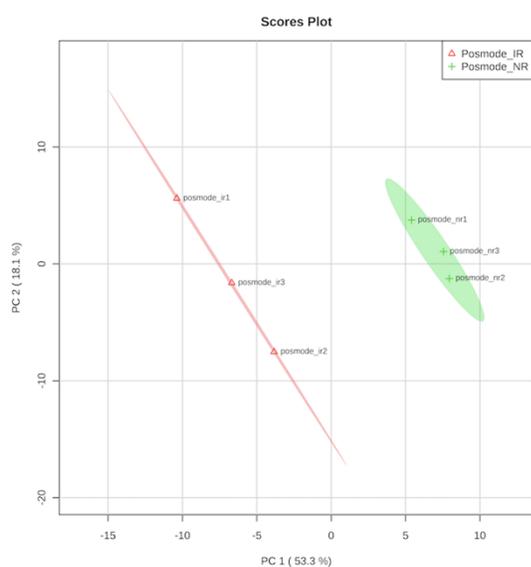




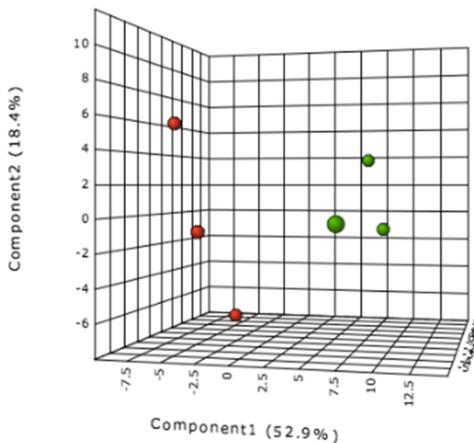
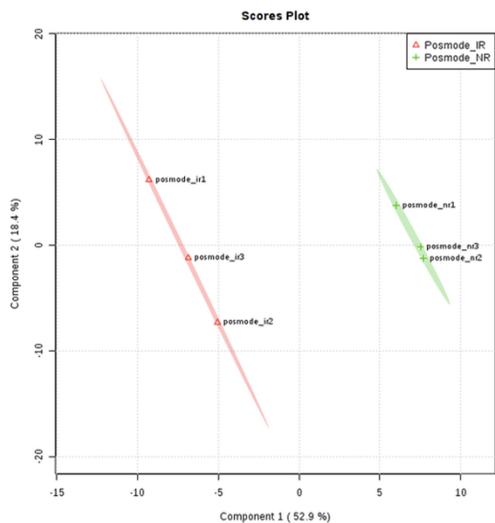
## Multivariate Analysis

- In this type of analysis, principal components are identified.
- Contributions from all the metabolites to each principal component are calculated, reducing >1000 values to one number for each component
- When this done in an unsupervised way, the group information is added after principal component analysis (PCA)
- Partial least squares discriminant analysis (PLS-DA) is a supervised procedure and group information is included in the analysis

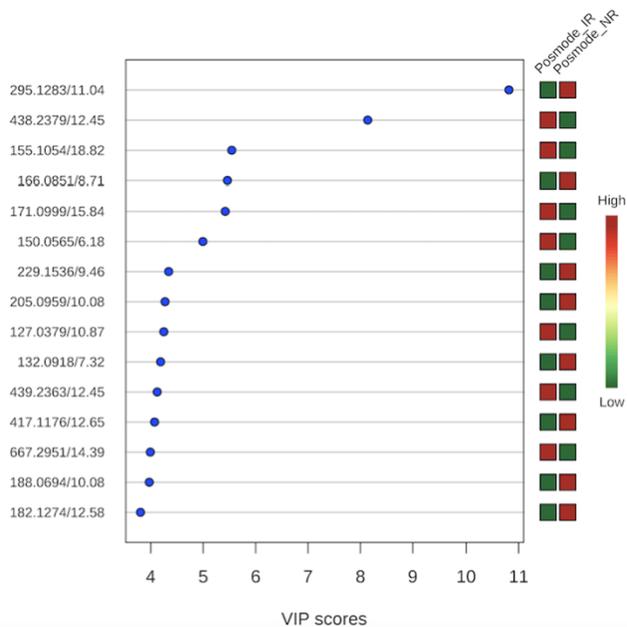
### 2D- and 3D-PCA plots



## 2D- and 3D-PLS-DA plots



## From download file – plsda\_vip.csv file



**Questions?**